STUDIES ON A NEW ALKALINE DEGRADATION PRODUCT OF JOSAMYCIN

Sir:

On the aglycon structure of macrolide antibiotics possessing an aldehyde function, Wo-ODWARD once suggested a 17-membered lactone structure with a -CHO side chain for magnamycin by degradation studies²⁾, and later revised it to a 16-membered structure with -CH2-CHO after NMR study of its degradation product.³⁾ For spiramycin, PAUL et al. proposed an 18-membered lactone structure with a -CHO side chain by degradation studies4), KUEHNE et al. amended it to a 17membered ring with a -CHO based on MS and NMR studies⁵⁾, and later OMURA et al. revised it to a 16-membered lactone with a -CH2-CHO by studying a derivative which was also obtained from leucomycin⁶). For leucomycin, OMURA et al. assigned a 16membered lactone structure with a -CH₂ -CHO side chain⁷⁾.

In the course of structure studies on josamycin¹⁾, we found a new interesting product which was obtained by mild alkaline treatment of this antibiotic.

When josamycin (I) was refluxed in ethanol with equimolar lithium hydroxide (LiOH \cdot H₂O) for three hours, a new product was formed

in good yield and was separated and purified by silica gel or alumina column chromatography. The purified substance (II) was obtained as a white amorphous powder, $[\alpha]_{\rm D}^{25}$ -20.7° (c 1, EtOH), $\lambda_{\rm max}^{\rm MeOH}$ 234.5 m μ (ε 25,900), and gave a crystalline thiosemicarbazone (III) as needles, mp 153~155°C, $\lambda_{\rm max}^{\rm MeOH}$ 232 m μ (ε 33,500) and 272 m μ (ε 28,400). II showed no antibacterial activity.

The elemental analysis of the crystalline thiosemicarbazone (III) showed excellent agreement for a formula C40H67NO18 ·NNHCSNH2 (Anal. Calcd. for C₄₁H₇₀N₄O₁₃S: C 57.32, H 8.21, N 6.52, S 3.73. Calcd. for C₄₁H₆₈N₄O₁₂S $(C_{41}H_{70}N_4O_{13}S-H_2O): C 58.55, H 8.15, N$ 6.66, S 3.81. Found: C 57.17, H 8.01, N 6.47, S 3.92). The mass spectrum of II gave the peak of highest mass at m/e 767, which could be assigned to the M⁺-18 peak derived from C40He7NO14 (MW 785) in consideration of the formula of III above-assigned. Thus the molecular weight of II was determined to be 785, which showed a decrease of 42 mass units from josamycin (I) (C42H69NO15, M^+ at m/e 827), suggesting hydrolytic removal of an acetyl from josamycin. In accordance with this, the NMR and IR spectra of II and its thiosemicarbazone (III) revealed loss of a signal at δ 2.28 (3 H, s, OCOCH₈) and a band at 1234 cm^{-1} (OCOCH₃ ν_{e-o}), indicating absence of O-acetyl originally found in I. Catalytic

Fig. 1. ORD and CD curves of josamycin (I) and its alkaline degradation product (II)



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Fig. 2. NMR spectrum of II (100 MHz, $CDCl_3+D_2O$)

Fig. 3. NMR spectrum of thiosemicarbazone of II (III) (100 MHz, $CD_3COCD_3+D_2O$)



hydrogenation of II over palladium showed the same amount of hydrogen absorption as that in I (two molar equivalents), denying formation of any new double bond by deacetylation. This was further confirmed by a ¹³C NMR spectrum of II giving signals for two ester carbonyl carbons (176.6 and 174.6 ppm) and four olefinic carbons (140.8, 135.9, 132.2 and 131.3 ppm), indicating a decrease of one ester carbonyl and no change in the number of double bonds compared with josamycin (I) (176.6, 174.5 and 173.6 ppm for ester carbonyls; 138.4, 135.3, 135.0 and 130.7 ppm for olefinic Thus II was shown to be a carbons). deacetylated josamycin.

Fig. 1 gives ORD and CD curves of II and josamycin (I). As seen in the figure, the ORD curves of II and I are almost symmetrical in the carbonyl region. II shows peaks at $319 \,\mathrm{m}\mu$ and $308 \,\mathrm{m}\mu$, while I gives troughs at 308 m μ and 298 m μ , showing positive and negative COTTON effect curves respectively. The CD curve of II shows positive maxima at $314 \text{ m}\mu$ (sh), $305 \text{ m}\mu$ ($\theta =$ +748) and 297 m μ (sh), and that of I gives negative maxima at 305 m μ (sh), 296 m μ ($\theta =$ -727) and 288 m μ ($\theta = -739$). These almost symmetrical ORD curves of II and I in the carbonyl region, suggested that II and I might have symmetrical partial structures in the vicinity of the aldehyde carbonyl group. This suggested the possibility of epimerization on the carbon atom adjacent to the aldehyde, indicating a 17-membered lactone aglycon in josamycin.

This epimer formation was also suggested by the 100 and 60 MHz NMR spectra of II and its thiosemicarbazone (III). The aldehyde proton of josamycin (I) gave a broad singlet signal at δ 9.64. II gave a doublet at δ 9.82 with a coupling constant of 2.0 Hz (cf. Fig. 2). In the thiosemicarbazone of II (III), the -CH=N- proton gave a well-resolved doublet at δ 7.60 (1 H, J=6.5 Hz). These J values were identical in 100 and 60 MHz spectra, and irradiation near 2.4 ppm caused collapse of the doublet at δ 7.60 into a singlet (Fig. 3). However, the ¹H FT-NMR spectrum of II on a Varian XL-100 NMR spectrometer kindly made by Dr H. NAGANAWA, Institute of Microbial Chemistry, Tokyo, showed that the aldehyde proton of II at δ 9.82 is a doubledoublet with coupling constants of 2.0 and 0.5 Hz. Decoupling studies showed that irradiation at δ 2.44 collapsed the broad doublet of the aldehyde proton into a broad singlet and irradiation at δ 2.14 caused sharpening of the doublet signal. Josamycin X-ray crystallography is now under study.

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